



Rapid Sterility Testing

Implementing Alternative Sterility Methods for Modern Pharmaceutical Manufacturing

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Agenda



Selecting a Rapid Method



Rapid ATP-Based testing for product sterility
(USP <73> and USP <1223> examples)

Poll Question #1

Which sterility testing method is primarily used at your organization?

- Traditional compendial sterility testing
- Alternative or rapid sterility testing
- A combination of both
- Not sure

Poll Question #2

What would be the most compelling reason for your organization to implement an alternative or rapid sterility test method?

- Faster time to product release
- Improved operational efficiency
- Enhanced data objectivity and automation
- Regulatory alignment and modernization
- Cost reduction
- To ask the lab for results (more quickly)

Poll Question #3

Where is your organization in implementing alternative or rapid sterility testing?

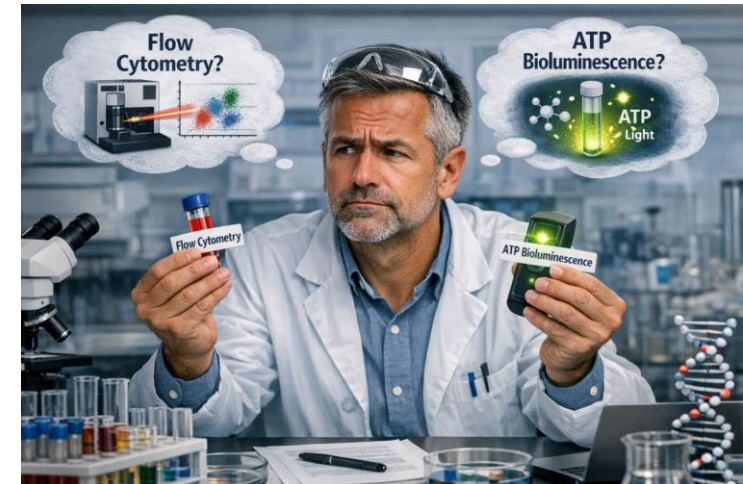
- Not currently considering it
- Exploring options
- In validation phase
- Fully implemented
- Previously evaluated but not adopted

A Brief History



Which Method is Right for My Use Case?

- Growth based
 - Electrochemical Measurement
 - CO₂ Detection
 - Biochemical/Carbohydrate Substrates
 - Digital Imaging / Auto-fluorescence
 - Laser Excitation
 - Selective Media
 - Head Space Pressure
 - Microcalorimetry
- Viability Based
 - Flow Cytometry
 - Laser Scanning Solid Phase Cytometry
 - Direct Epifluorescence Filter Microscopy
- Nucleic Acid Amplification
 - PCR
 - RT PCR
 - Ribotyping
 - Gene Sequencing
- Cellular Component Based
 - ATP Bioluminescence
 - Fatty Acid Profiling
 - MALDI-TOF
 - SELDI-TOF
 - FT-IR
 - Endotoxin Detection
- Optical Spectroscopy
 - Light Scattering/Intrinsic Fluorescence
 - Raman Spectroscopy
- Micro-Electrical-Mechanical Systems (MEMS)
 - Microfluidics
 - Microarrays



Which Method is Right for My Use Case?

Technology	LOD (CFU)	Time to Result	Sample Size Range (mL)
Chapter (71) (for comparative purposes only)	Theoretical LOD of 1–3 CFU based on a Poisson distribution	14 days	Refer to (71)
Adenosine triphosphate (ATP) bioluminescence	1–10	2–7 days (including pre-enrichment)	0.1–1000
Nucleic acid-based amplification	10–100	2–4 h	0.2–2
Respiration	1–10	Overnight to 7 days	Up to 10 per bottle
Solid phase cytometry	1–10	2–8 h	1–1000

Chart from USP [\(1071\)](#)

Consult with your Regulators!



Work with regulatory agencies/notified bodies on proposed AMM validation plan



Define product requirements based on critical factors (filing info, 510(k) vs PMA, MDR status, etc.)



Understand regional requirements, as these may vary

User Requirement Specifications

- A rapid result time
- Microorganism Detection:
 - Low quantity
 - Wide range
 - Specific organisms of concern
 - Investigations
- Samples:
 - Quantity
 - Batches
 - Handling



- Instrument:
 - Aseptic test material handling
 - Availability of instruments and reagents
 - Reference material and Controls
 - Ease of use/simplicity of test and data interpretation
 - Low rates of false positive and false negative results
 - Manual vs automatic preparation methods
 - Automation and automated continuous/periodic monitoring
- Improved patient safety
- Robustness
- Reliability



ATP Bioluminescence: Pros and Cons

Advantages

- Broad equivalency to compendial methods
- Early detection of microbial growth
- Isolation of organism for identification
- Easily fits into current workflow
- Objective Results
- Vendors and reagents widely available

Disadvantages

- Inability to detect organisms that do not grow under conditions utilized
- Products with significant ATP background that cannot be reduced
- Increased labor time and cost of testing

Rapid Product Sterility: AMM Support in Standards

- USP <1223>: Validation of Alternative Microbiological Methods
- Ph. Eur 5.1.6: Alternative Methods for Control of Microbiological Quality
- PDA Technical Report 33: Evaluation, Validation and Implementation of Alternative and Rapid Microbiological Methods (New revision effective soon)

Revised or newly official 01 Aug 2025

- USP <1071>: Rapid Microbial Tests for Release of Sterile Short-Life Products: A Risk-Based Approach
- USP <72>: Respiration-Based Microbiological Methods for the Detection of Contamination in Short-Life Products
- USP <73>: ATP Bioluminescence-Based Microbiological Methods for the Detection of Contamination in Short-Life Products

Others:

- USP <74>: Solid Phase Cytometry-Based Microbiological Method for the Detection of Contamination in Clear Aqueous Solutions (published in PF51(3) in May 2025)
- USP <75>: Nucleic Acid Amplification Method for the Detection of Contamination in Short-Life Products (under development)

Requirements for USP <73>

- **Short-life products:** Products with a short-shelf life or with a short time between manufacturing and administration and for which a sterility test requiring a 14-day incubation is not suitable.
- Primary validation of the method per USP <1071>
 - Does **not** require equivalence/comparability to USP <71>
- Should include standard method suitability organisms AND a **selection of organisms relevant to product/manufacturing process.**
 - If relevant, slow-growing organisms and local isolates may be included
- Inoculum level for method suitability is ≤ 10 CFU
- Perform sample interference study
- Incubation time determined via calculation plus **safety factor**

Generation Time Calculation for USP <73>

GENERATION TIME

- Generation time = time required for microbial cell number to double

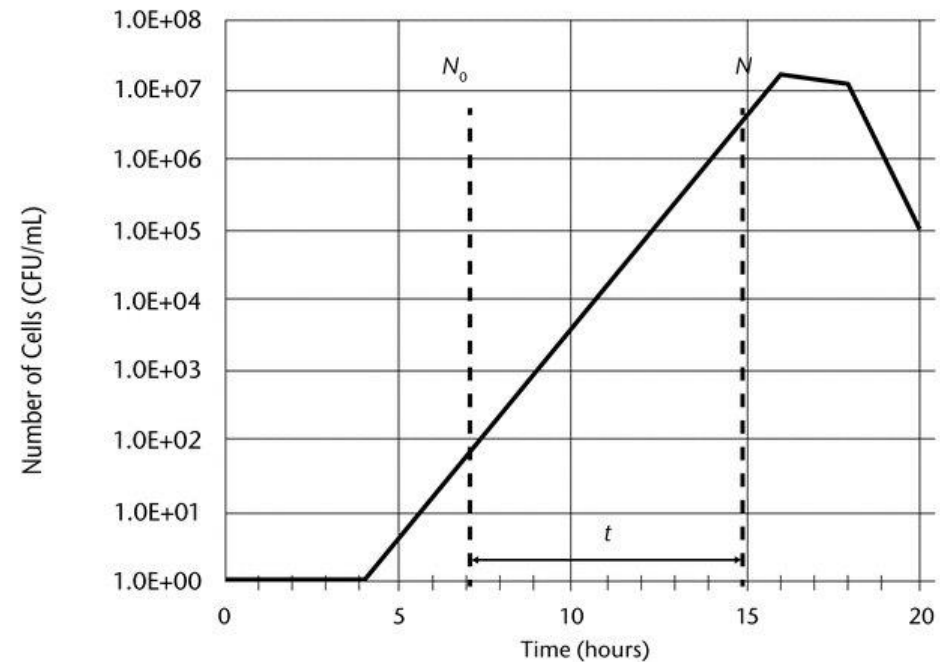
$$G = \frac{t}{3.3 \times \log_{10}\left(\frac{N}{N_0}\right)}$$

t = time interval (h)

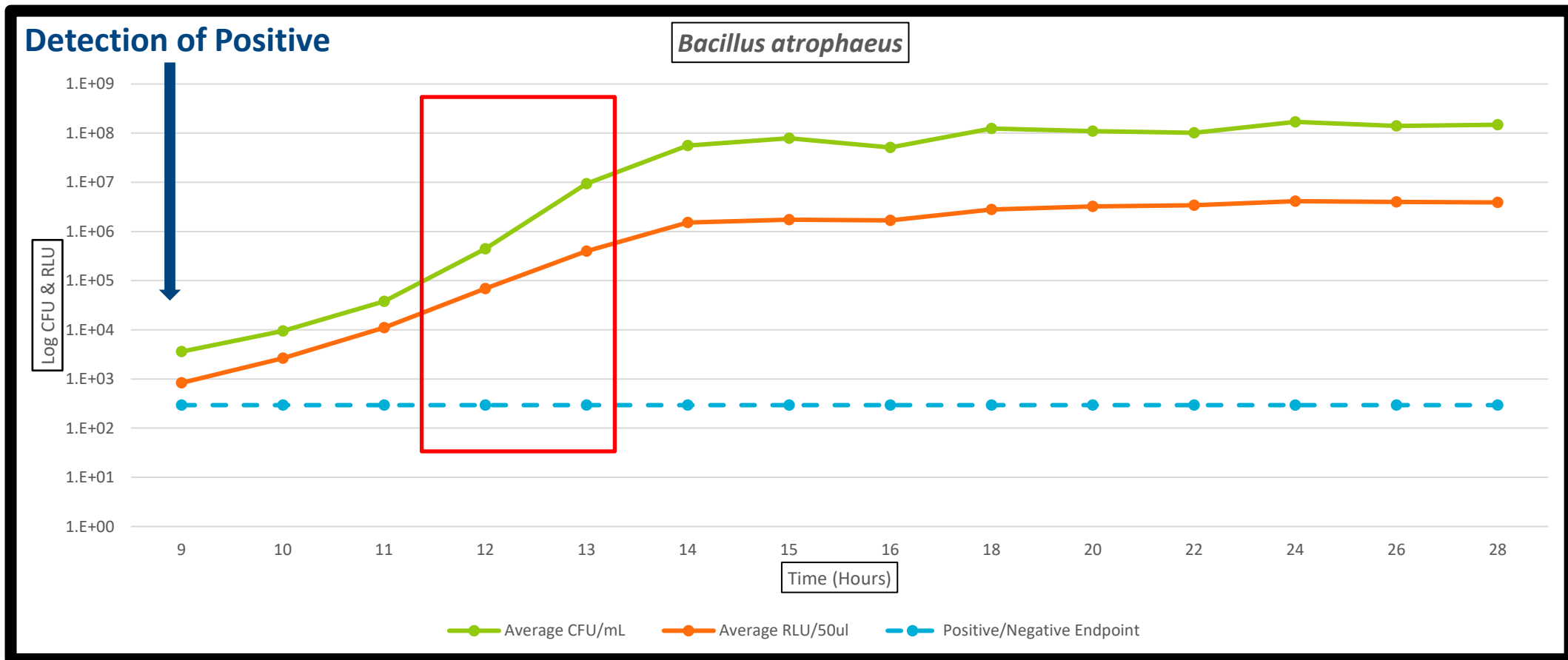
N = number of cells/CFU at the end of the time interval

N_0 = number of cells/CFU at the beginning of the time interval

EXAMPLE



Generation Time Example



Incubation Time Calculation for USP <73>

USP <73> CALCULATION EXAMPLE

- Slowest organism time to detection of 48 hours
- Generation time of 1 hour

$$T = t_{ttd} + \log_2(10) \times G$$

$$T = 48h + 3.32h \times 1$$

$$T = 51.3h \text{ or } 2.1 \text{ days}$$

T = incubation time for microbial detection in the product to be examined (h)

t_{ttd} = longest time to detection in the method suitability test (h)

G = generation time slowest growing organism (h)

BACILLUS ATROPHAEUS CALCULATION EXAMPLE

- Time to Detection (+): 9 hours
- Generation (G) Time: 0.29 hours
- $T = 9h + \log_2(10) \times G$
- $T = 9h + (3.32h \times 0.29)$
- $T = 9.96h$ (approximately a 10h minimum incubation time)

⟨1223⟩ VALIDATION OF ALTERNATIVE MICROBIOLOGICAL METHODS

Qualitative tests must meet certain validation parameters

Examples of qualitative tests: 1) sterility test and 2) test for absence of specified microorganisms

Validation parameters that are required include the following

Specificity

Limit of Detection

Robustness

Repeatability

Ruggedness

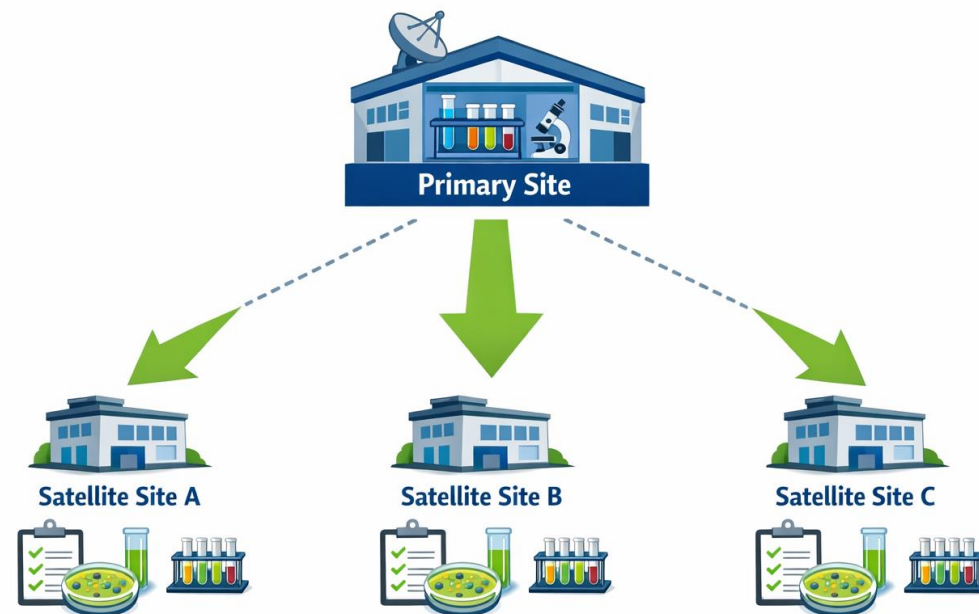
Equivalency

Suitability

Requirements	<73>	<1223>
Method Suitability:		
USP <71> Organisms	✓	✓
Environmental Isolates (3)	✓	✓
≤ 10 CFU	✓	
≤ 100 CFU		✓
False Positive/Negative:		
Sample Interference	✓	✓

Multi-Site Implementation

- Are there any differences in the sites?
 - Temp, rH, altitude
 - Microflora
 - Data recording
- Keep IQ/OQ consistent
- Reduced PQ
 - Ensure all site-specific materials are assessed





The Rules

- There are 5 samples, note which are positive and which are negative
- The samples were processed through the ATP Bioluminescence reader to determine a Relative Light Unit (RLU) value which correlates to ATP activity in the sample
- If the RLU value is 3 times greater than the negative control, it is considered positive

The Results

Sample #	Visual Result	RLU Result	Sample Type
1			
2			
3			
4			
5			

An Even Simpler Approach

Requirements	Do it Yourself	Contract Lab
IQ	✓	Already established
OQ	✓	Already established
PQ	✓	Already established
<71> Equivalence Testing	✓	Already established
Suitability	✓	✓

Conclusions

Consult with your regulators early and have a plan

Know your user requirements

Rapid sterility is an impactful way to shorten release times and improve patient safety

Questions?





Useful Links

Sotera Health Academy:



White Paper:



LinkedIn:

